

GSMR

Generalised
Summary-data-
based Mendelian
Randomisation

GCTA

SMR

GSMR

OSCA

GCTB

Program in CTG

CTG forum

Overview

The **gsmr** R-
package

implements
the GSMD
(Generalised
Summary-
data-based
Mendelian
Randomisation)
method to

test for
putative
causal
association
between a
risk factor and
a disease
using

summary-
level data
from genome-
wide
association
studies
(GWAS) (Zhu
et al. 2018)

Nat.

Commun.).

The R

package is

developed by

Zhihong Zhu,

Zhili Zheng,

Futao Zhang

and Jian Yang

at Institute for

Molecular

Bioscience,

the University

of

Queensland.

Bug reports or

questions:

jian.yang@uq.edu

Note: The

GSMR

method has

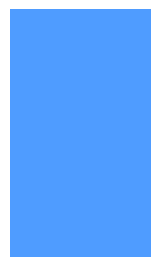
also been

implemented

in the GCTA

software

(GCTA-
GSMR)



Citation

Zhu, Z. et

al. (2018)

Causal

associations
between risk
factors and
common
diseases
inferred from
GWAS
summary

data. Nat.

Commun. 9,

224

(<https://www.nature.com/doi/10.1038/017-02317-2>).

Source

code

[gsmr_1.0.9.tar.gz](#)

Note: We
included a
new HEIDI-
outlier method
(as part of the
GSMR

analysis) in
gsmr v1.0.7.
However, the
new HEIDI-
outlier method
is currently
under
development

and subject to
changes
during the
method
development.

From the

GSMR R

package (\geq)

version 1.0.8),
we changed
the default
back to the
original HEIDI-
outlier method
described in
Zhu et

al. (2018

Nature

Communications)

and added a

temporary

flag

(‘gsmr2_beta’)

to test the

new method.

The command

to use this

flag can be

found in the

tutorial below.

The new

HEIDI-outlier

method in
gsmr (\geq
version 1.0.8)
has been
tested by
extensive
simulations
and real data

analyses. We
will make a
formal release
in our next
GSMR paper.

Sample data
are available
in

[test_data.zip](#).

This

document has

been

integrated in

the gsmr R-

package, we

can check it

by the
standard
command “?
function_name”
in R.

Installation

The **gsmr**

requires R \geq
2.15, you can
install it in R
by:

```
# gsmr req  
quires the  
R-package(  
s)  
install.pa
```

```
packages(c('
survey')));
# install
gsmr
install.pa
ckages("ht
tp://cnsge
nomics.com
/software/
gsmr/stati
c/gsmr_1.0
0 + on all
```

```
1.9.tar.gz  
, repos=NUL  
L, type="so  
urce")
```



Update log

V1.0.9

(gmr_1.0.9.tar.gz

PDF, 18

Jun. 2019):

Change the

flag

‘gsmr_beta’

to

‘gsmr2_beta’.

V1.0.8

(gmr_1.0.8.tar.gz

PDF, 21

Jan. 2019):

Added a flag

‘gsmr_beta’

to use a

testing

version of the

HEIDI-outlier
method.

V1.0.7

([gmr_1.0.7.tar.gz](#)

[PDF](#), 9

Oct. 2018):

Added a

multi-SNP-

based HEIDI-
outlier test in
the HEIDI-
outlier
analysis.

V1.0.6

[\(gmr_1.0.6.tar.gz](#)

[PDF, 23](#)

Jan. 2018):

Added a

function to

remove SNPs

in high LD.

V1.0.5

([gmr_1.0.5.tar.gz](#)

[PDF](#), 13

Dec. 2017):
Improved the
approximation
of the
sampling
covariance
matrix.

V1.0.4

([gsmr_1.0.4.tar.gz](https://github.com/chr17huggins/gsmr_1.0.4.tar.gz);

PDF, 6

Nov. 2017):

Added the bi-

directional

GSMR

analysis. The

HEIDI-outlier

analysis has
been
integrated in
the GSMR
analysis by
default.

V1.0.3

([gsmr_1.0.3.tar.gz](#))

PDF, 12

Oct. 2017):

Added more
example data.

Removed the
initial versions
(8 Nov 2016).

Tutorial

The GSMMR
analysis only
requires
summary-
level data
from GWAS.

Here is an
example,
where the risk
factor (x) is
LDL
cholesterol
(LDL-c) and
the disease (y)

is coronary
artery disease
(CAD). GWAS
summary data
for both LDL-
c and CAD
are available
in the public

domain

(Global Lipids

Genetics

Consortium et

al. 2013,

Nature

Genetics;

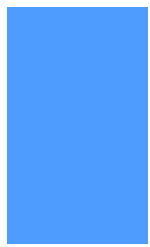
Nikpay, M. et

al. 2015,
Nature
Genetics).



1.

**Prepare
data for
GSMR**



analysis

1.1 Load the GWAS summary data

```
library("gsmr")  
data("gsmr")  
head(gsmr_data)
```

```
##
```

```
SNP a1 a2
```

```
a1 freq
```


at_freq

bzx bzx_se

bzx_pval

bzx_n

bzy

1 rs109

03129 A

G 0.450019

47 -0.0328

0.0037 3.0

30e-17 169

920.0 0.0

08038

2 rs127

48152 T

C 0.080877

58 0.0499

0.0066 3.2

09e-12 172

987.5 0.0

13671

3 rs112

06508 A

G 0.143969

88 0.0434

0.0055 2.2

56e-14 172

239.0 0.0

30222

4 rs112

06510 C

T 0.191289

11 -0.0831

0.0050 2.3

80e-53 172

012 0 0 0

012.0 -0.0

74519

5 rs107

88994 T

C 0.183954

30 0.0687

0.0049 8.8

67e-41 172

941.9 0.0

38267

6 rs5

29787 G

```
C 0.197130
99 -0.0553
0.0052 8.7
46e-24 161
969.0 0.0
01707
##          bz
y_se      b
zy_pval   b
zy_n
## 1 0.009
2442 0.384
```

5651000 18

4305

2 0.018

5515 0.461

1690000 18

4305

3 0.014

1781 0.033

0400000 18

4305

4 0.013

0400000 18

3438 0.000

0000234 18

4305

5 0.011

8752 0.001

2711000 18

4305

6 0.013

5491 0.899

7431000 18

4305

```
dim(gsmr_data)
```

```
## [1] 188  
12
```

This is the
input format

for the GSMMR
analysis. In
this data set,
there are 188
near-
independent
SNPs
associated

with LDL-c at
a genome-
wide
significance
level (i.e. $p < 5e-8$).

- SNP: the genetic

instrument

- a1: effect allele
- a2: the other allele
- a1_freq: frequency of a1
- bzx: the

effect size
of a_1 on
risk factor

- bzx_se :
standard
error of bzx
- bzx_pval : p
value for
 bzx

- `bzx_n`: per-SNP sample size of GWAS for the risk factor
- `bzy`: the effect size

of a_1 on
disease

- bzy_se :
standard
error of bzy
- bzy_pval : p
value for
 bzy
- bzy_n : per-

SNP
sample
size of
GWAS for
the disease

**1.2 Estimate
the LD
correlation
matrix**

```
# Save the genetic variants and effect alleles in a text file using R  
write.table(gsmr_data[,c(1,2)], "gsmr_ex
```



```
ample_snps
.allele",
col.names=
F, row.names=
F, quote
=F)
# Extract
the genoty
pe data fr
om a GWAS
dataset us
ing GCTA
```

```
gcta64 --b  
file gsmr_  
example --  
extract gs  
mr_example  
_snps.alle  
le --updat  
e-ref-alle  
le gsmr_ex  
ample_snps  
.allele --
```

```
recode --o  
ut gsmr_ex  
ample
```

Note: the two
steps above
guarantee
that the LD
correlations

are calculated based on the effect alleles for the SNP effects.

```
# Estimate  
LD correla  
tion matri  
x using R
```

```
x using R
snpcoeff_
id = scan(
"gsnr_exam
ple.xmat.g
z", what="
", nlines=
1)

snpcoeff
= read.tab
le("gsnr_e
xample.xma
```

```
t.gz", header=F, skip=2)
```

```
# Match the SNP genotype data with the summary data  
cnp_id = D
```

```
snp_id = r  
reduce(intersect, list(gsmr_data  
a$SNP, snp  
_coeff_id)  
)  
gsmr_data  
= gsmr_data  
a[match(snp  
_id, gsmr  
_data$SNP)
```

```
, ]  
snp_order  
= match(snp_  
p_id, snp_  
coeff_id)  
snp_coeff_  
id = snp_c  
oeff_id[snp_  
p_order]  
snp_coeff  
= snp_coef  
f[, snp_or
```



```
der]
```

```
# Calculate the LD correlation matrix
```

```
ldrho = cor(snp_coef)
```

```
# Check th
```

e size of
the correl
ation matr
ix and dou
ble-check
if the ord
er of the
SNPs in th
e LD corre
lation mat
rix is con
sistent wi

th that in
the GWAS s
ummary dat
a

```
colnames(ldrho) = rownames(ldrho) = snp_  
coeff_id
```

```
dim(ldrho)
```

```
## [1] 188
```

```
188
```

```
# Show the
first 5 ro
ws and col
umns of th
e matrix
ldrho[1:5,
1:5]
```

```
##
```

```
rs10003120
```

rs10903129

rs12748152

rs11206508

rs11206510

rs10788994

rs10903

129 1.000

000000 -0.

0045378845

0.00806662

1 -0.01372

112 -0.023

4447102

rs12748

152 -0.004

537884 1.

0000000000

-0.0066871

81 0.0044

5927 0.00

03629201

rs11206

508 0.008

066621 -0.

0066871806

1.0000000000

0 -0.21125

757 0.051

2593434

rs11206

510 -0.013

721120 0.

0044592696

-0.2112575

67 1.00000

0000 0 10

0000 -0.10

42706205

rs10788

994 -0.023

444710 0.

0003629201

0.05125934

3 -0.18427

062 1.000

0000000

Note: all the analyses implemented in this R-package only require the summary data (e.g. “gsmr_data”)

and the LD
correlation
matrix
(e.g. “ldrho”)
listed above.

2.

Standardiza

This is an
optional
process. If the
risk factor
was not
standardised
in GWAS, the
effect sizes

can be scaled
using the
method
below. Note
that this
process
requires allele
frequencies,

z-statistics
and sample
size. After
scaling, b_{zx} is
interpreted as
the per-allele
effect of a
SNP on the

exposure in
standard
deviation
units.

```
snpfreq =  
gsmr_data$a1_freq  
# allele frequencies
```

```
of the SNP  
s  
bzx = gsmr  
_data$bzx  
# effects  
of the ins  
truments o  
n risk fac  
tor  
bzx_se = g  
smr_data$b  
zx_co
```



```
zx_se  
# standard  
errors of  
bzx  
bzx_n = gs  
mr_data$bz  
x_n  
# GWAS sam  
ple size f  
or the ris  
k factor  
std_zx = s
```

```
td_effect(  
  snpfreq, b  
  zx, bzx_se  
  , bzx_n)  
# perform  
standardis  
ation  
gsmr_data$  
std_bzx =  
std_zx$b  
# standard  
ized bzx
```

```
gsmr_data$  
std_bzx_se  
= std_zx$s  
e # sta  
ndardized  
bzx_se  
head(gsmr_  
data)
```

```
##
```

```
SNP a1 a2
a1_freq
bzx bzx_se
bzx_pval
bzx_n
bzy
## 1 rs109
03129 A
G 0.450019
47 -0.0328
0.0037 3.0
30e-17 169
```

920.0 0.0

08038

2 rs127

48152 T

C 0.080877

58 0.0499

0.0066 3.2

09e-12 172

987.5 0.0

13671

3 rs112

06500 A

06508 A

G 0.143969

88 0.0434

0.0055 2.2

56e-14 172

239.0 0.0

30222

4 rs112

06510 C

T 0.191289

11 -0.0831

0.0050 2.3

```
80e-53  172
812.0   -0.0
74519
## 5   rs107
88994   T
C  0.183954
30     0.0687
0.0049  8.8
67e-41  172
941.9   0.0
38267
## 6     rs5
```

29787 G

C 0.197130

99 -0.0553

0.0052 8.7

46e-24 161

969.0 0.0

01707

bz

y_se b

zy_pval b

zy_n s

td_bzx st

d_bzx_se

1 0.009

2442 0.384

5651000 18

4305 -0.03

055942 0.0

03447252

2 0.018

5515 0.461

1690000 18

4305 0.04

713698 0.0

06234550

3 0.014

1781 0.033

0400000 18

4305 0.03

829018 0.0

04852442

4 0.013

3438 0.000

0000234 18

4305 -0 07

4305 0.07

181919 0.0

04321251

5 0.011

8752 0.001

2711000 18

4305 0.06

149455 0.0

04386074

6 0.013

5491 0.899

7431000 18

4305 -0.04

695042 0.0

04414868

3.

GSMR

analysis

This is the

main analysis
of this R-
package. It
uses SNPs
associated
with the risk
factor (e.g. at
 $p < 5e-8$) as

the
instruments to
test for
putative
causal effect
of the risk
factor on the
disease. The

analysis

involves a

step that uses

the **HEIDI-**

outlier

approach to

remove SNPs

that have

effects on
both the risk
factor and the
disease
because of
pleiotropy.

```
bzx = gsmr  
_data$std_
```



```
bzx      # S
NP effects
on the risk factor
bzx_se = gsmr_data$
td_bzx_se
# standard
errors of
bzx
bzx_pval =
gsmr_data$
```

```
bzx_pval
# p-values
for bzx
bzy = gsmr
_data$bzy
# SNP effects on the
disease
bzy_se = g
smr_data$b
zy_se      #
```

```
Standard e  
rrors of b  
zy  
bzy_pval =  
gsmr_data$  
bzy_pval  
# p-values  
for bzy  
n_ref = 77  
03      # Sa  
mple size  
of the ref
```

reference sam
ple

gwas_thres

h = $5e-8$

GWAS thr

eshold to

select SNP

s as the i

nstruments

for the GS

MR analysi

s

```
single_snp  
_heidi_thr  
esh = 0.01  
# p-value  
threshold  
for single  
-SNP-based  
HEIDI-outl  
ier analys  
is  
multi_snp_
```

```
heidi_thre  
sh = 0.01  
# p-value  
threshold  
for multi-  
SNP-based  
HEIDI-outl  
ier analys  
is  
nsnps_thre  
sh = 10  
# the mini
```

minimum number
of instruments
required for the
GSMR analysis

heidi_outlier_flag =
T # flag for HEIDI
I-outlier
analysis

and cys13

ld_r2_thre

sh = 0.05

LD r2 th

reshold to

remove SNP

s in high

LD

ld_fdr_thr

esh = 0.05

FDR thre

shold to r

remove the
chance cor
relations
between th
e SNP inst
ruments
gsmr2_beta
= 0 #
0 – the or
iginal HEI
DI-outlier
method; 1

– the new
HEIDI-outlier method
that is currently under development

```
gsmr_results = gsmr(  
    bzx, bzx_se,  
    bzx_pval,  
    bzy, bz
```

l, bzy, bz
y_se, bzy_
pval, ldrh
o, snp_coe
ff_id, n_r
ef, heidi_
outlier_fl
ag, gwas_t
hresh, sin
gle_snp_he
idi_thresh
, multi_sn

```
p_heidi_th  
resh, nsnp  
s_thresh,  
ld_r2_thre  
sh, ld_fdr  
_thresh, g  
smr2_beta)  
# GSMM ana  
lysis  
filtered_i  
ndex=gsmr_  
results$us
```

```
ed_index  
cat("The e  
stimated e  
ffect of t  
he exposur  
e on outco  
me: ", gsmr  
_results$b  
xy)
```

```
## The estimated effect of the exposure on outcome:  
0.4322395
```

```
cat("Standard  
error  
of bxy: ",  
gsmr_results$  
bxy_se)
```

```
## Standard  
error of  
bxy: 0.02  
210985
```



```
cat("P-value for bxy  
: ", gsmr_  
results$bxy_  
pval)
```

P-value
for bxy:
4.15454e-8
5

```
cat("Indexes of the  
SNPs used  
in the GSM  
R analysis  
: ", gsmr_  
results$us  
ed_index[1  
:5], "..."  
)
```

```
## Indexes  
of the SNP  
s used in  
the GSMR a  
nalysis:
```

```
1 2 3 5 6
```

```
...
```

```
cat("Number of SNPs  
with missing  
estimates in the  
summary data:  
", length(gsmr_results$na_snp))
```

```
## Number  
of SNPs wi  
th missing  
estimates  
in the sum  
mary data:  
0
```

```
cat("Number of non-significant  
SNPs: ", length(gsmr_results$weak_snps))
```

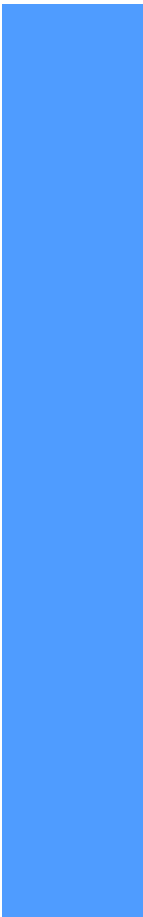
Number
of non-sig
nificant S
NPs: 39


```
cat("Number of SNPs  
in high LD  
( LD rsq >  
", ld_r2_t  
hresh, "):  
", length(  
gsmr_result  
ts$linkage  
_snps))
```

```
## Number  
of SNPs in  
high LD (  
LD rsq > 0  
.05 ): 5
```

```
cat("Number of pleio  
tropic outliers: ",  
length(gsm  
r_results$  
pleio_snps  
))
```

```
## Number  
of pleiotr  
opic outli  
ers: 9
```



4. Bi- directional GSMR

analysis

The script
below runs bi-
directional
GSMR
analyses,
i.e. a forward-
GSMR

analysis as
described
above and a
reverse-
GSMR
analysis that
uses SNPs
associated

with the
disease
(e.g. at $p < 5e-8$) as the
instruments to
test for
putative
causal effect

of the disease
on risk factor.

```
gsmr_results = bigsmr(bzx, bzx_se, bzx_pval, bzy, bzy_se, bzy_pval, ld_rho, cnp_c
```



```
rno, snp_c  
coeff_id, n  
_ref, heid  
i_outlier_  
flag, gwas  
_thresh, s  
ingle_snp_  
heidi_thre  
sh, multi_  
snp_heidi_  
thresh, ns  
nps_thresh
```

```
, ld_r2_th  
resh, ld_f  
dr_thresh,  
gsmr2_beta  
) # GSM  
R analysis
```

```
cat("Effec  
t of risk  
factor on  
disease: "  
, gsmr resu
```

```
lts$forward_bxy)
```

```
## Effect  
of risk fa  
ctor on di  
sease: 0.  
4322395
```

```
cat("Standard error  
of bxy in  
the forward-GSMR ana-  
lysis: ", gsmr_result  
s$forward_bxy_se)
```

```
## Standard error of  
bxy in the  
forward-GS  
MR analysis:  
0.0221  
0985
```

```
cat("P-value of bxy  
in the forward-GSMR  
analysis:  
", gsmr_results$forward_bxy_pval)
```

P-value
of bxy in
the forward-GSMR ana-
lysis: 4.
15454e-85

```
cat("Effect  
of disease  
on risk  
factor: ",  
gsmr_results$reverse  
_bxy)
```



```
## Effect  
of disease  
on risk fa  
ctor:    -0.  
02739421
```

```
cat("Standard error  
of bxy in  
the reverse  
e-GSMR ana  
lysis: ", g  
smr_result  
s$reverse_  
bxy_se)
```

```
## Standard error of  
bxy in the  
reverse-GS  
MR analysis:  
0.0095  
51025
```

```
cat("P-value of bxy  
in the reverse-GSMR  
analysis:  
", gsmr_results$reverse_bxy_pval)
```

```
## P-value  
of bxy in  
the revers  
e-GSMR ana  
lysis: 0.  
004128198
```

5.

Visualization

```
effect_col  
= colors()  
[75]  
vals = c(b  
zx[filtere  
d_index]-b  
zx_se[filt  
ered_index  
, bzx[fil  
tered_inde  
x]+bzx se[
```

```
filtered_index])
xmin = min
(vals); xmax
= max(vals)
vals = c(b
zy[filtere
d_index]-b
zy_se[filt
ered_index
1 b-v[f:1
```

```
], bzy[filtered_index]+bzy_selected_index])  
ymin = min(vals); ymax = max(vals)  
par(mar=c(5,5,4,2))  
plot(bzx[f
```



```
filtered_in  
dex], bzy[  
filtered_i  
ndex], pch  
=20, cex=0  
.8, bty="n  
", cex.axi  
s=1.1, cex  
.lab=1.2,  
co  
l=effect_c  
ol. xlim=c
```

```
(xmin, xmax), ylim=c  
(ymin, ymax),
```

```
      xl  
ab=expression(LDL~cholesterol~  
(italic(b[  
zx]))),
```

```
      yl
```

```
ab=expression(Corona  
ry~artery~  
disease~(i  
talic(b[zy  
]))))
```

```
abline(0,  
gsmr_results$forward  
_bxy, lwd=  
1.5, lty=2  
, col="dim
```

```
grey" )
```

```
nsnps = length(bzx[filtered_index])
```

```
for( i in 1:nsnps )
```

```
{
```

```
    # x ax
```

```
is
```

```
    xstart
```

```

xstart
= bzx[filter
ered_index
[i]] - bzx
_se[filter
ed_index[i
]]; xend =
bzx[filter
ed_index[i
]] + bzx_s
e[filtered
_index[i]]

```

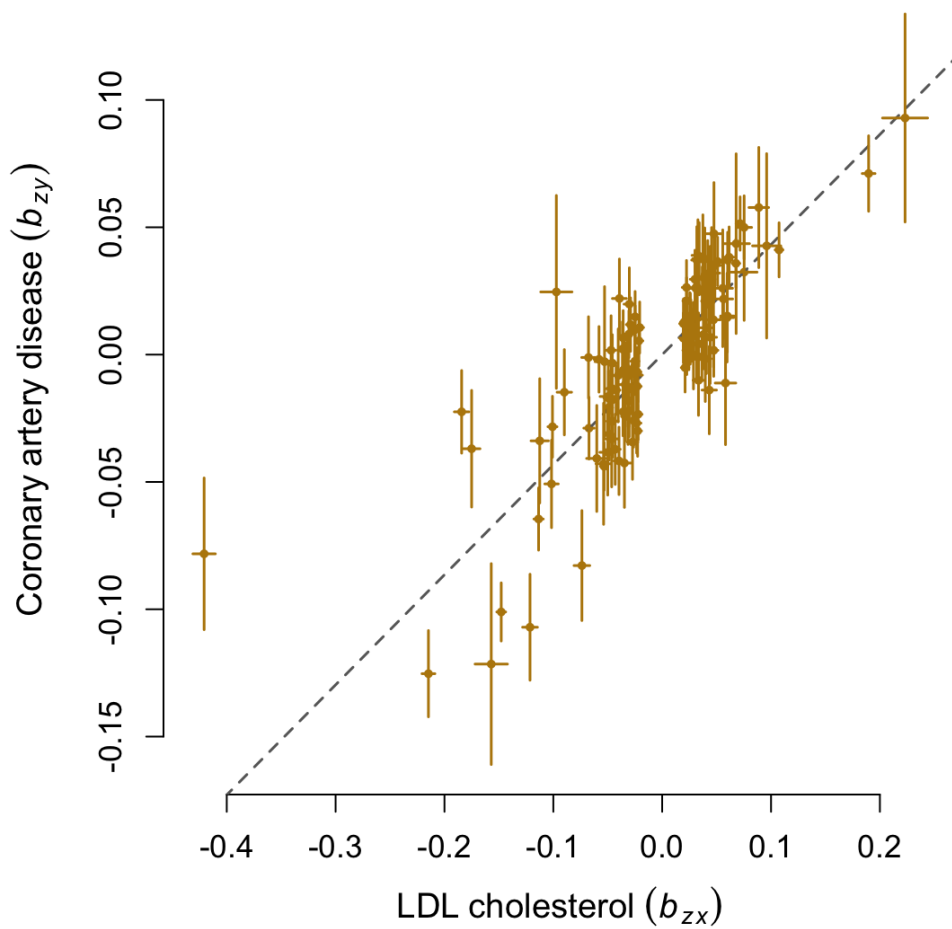
```
        ystart  
= bzy[filter  
ered_index  
[i]]; yend  
= bzy[filter  
ered_index  
[i]]  
  
        segmen  
ts(xstart,  
ystart, xe  
nd, yend,  
lwd=1.5, c
```

```
ol=effect_  
col)  
    # y ax  
is  
    xstart  
= bzx[filter  
ered_index  
[i]]; xend  
= bzx[filter  
ered_index  
[i]]  
    vstart
```

```
        ystart  
= bzy[filter  
ed_index  
[i]] - bzy  
_se[filter  
ed_index[i  
]]; yend =  
bzy[filter  
ed_index[i  
]] + bzy_s  
e[filtered  
_index[i]]
```



```
segments(xstart,  
ystart, xend,  
yend,  
lwd=1.5, col=effect_  
col)  
}
```




Package

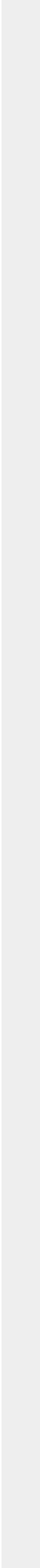
Document



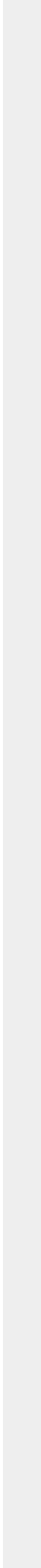
bi_gsmr



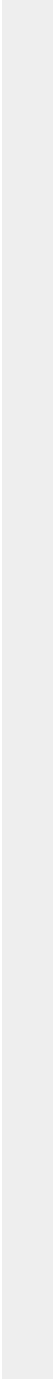
Bi-
directional
GSMR
analysis is
composed



of a
forward-
GSMR
analysis
and a
reverse-
GSMR
analysis
that uses



SNPs
associated
with the
disease
(e.g. at $< 5e-8$) as
the
instruments
to test for



putative
causal
effect of
the disease
on the risk
factor.

Usage

```
bi_gsmr(bzx
, bzx_se, b
zx_pval, bz
y, bzy_se,
bzy_pval, l
drho, snpid
, heidi_out
lier_flag=T
, gwas_thre
sh=5e-8, si
ngle_snp_he
```

```
idi_thresh=  
0.01, multi  
_snp_heidi_  
thresh=0.01  
, nsnp_s_thr  
esh=10, ld_  
r2_thresh=0  
.05, ld_fdr  
_thresh=0.0  
5, gsmr2_be  
ta=0)
```


Arguments

`bxz`

bzx_se

bzx_pval

bzy

bzy_se

bzy_pvał

ldrho

snpid

n_ref

heidi_outlier fla

gwas_thresh

single_snp_heidi.

multi_snp_heidi_.

nsnps_thresh

ld_r2_thresh

ld_fdr_thresh

gsmr2_beta

Value

Estimate of

causative
effect of risk
factor on
disease
(forward_bxy),
the
corresponding
standard error

(forward_bxy_se),

p-value

(forward_bxy_pvalue

and SNP

index

(forward_index),

and estimate

of causative

effect of
disease on
risk factor
(reverse_bxy),
the
corresponding
standard error
(reverse_bxy_se),

p-value

(reverse_bxy_pva

SNP index

(reverse_index),

SNPs with

missing

values, with

non-

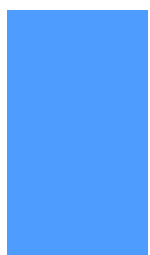
significant p-values and those in LD.

Examples

```
data("gsmr")  
gsmr_result = bi_gsmr  
# / ... /
```

```
r(gsmr_data  
a$bzx, gsm  
r_data$bzx  
_se, gsmr_  
data$bzx_p  
val, gsmr_  
data$bzy,  
gsmr_data$  
bzy_se, gs  
mr_data$bz  
y_pval, ld  
rho, gsmr_
```

```
data$SNP,  
n_ref, T,  
5e-8, 0.01  
, 0.01, 10  
, 0.05, 0.  
05, 0)
```



gsmr



GSMR

(Generalised

Summary-

data-based

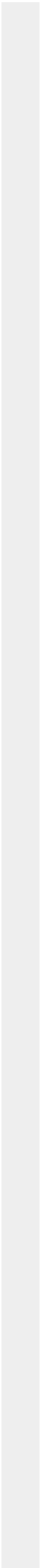
Mendelian

Randomisation

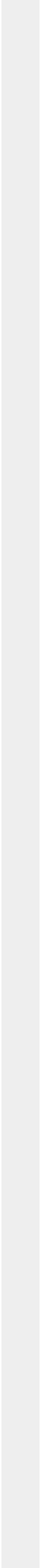
is a flexible

and

powerful



approach
that utilises
multiple
genetic
instruments
to test for
causal
association
between a



risk factor
and
disease
using
summary-
level data
from
independent
genome-

wide
association
studies.

Usage

```
gsmr(bzx, b  
zx_se, bzx_  
pval, bzy,  
bzy_se, ldr
```

```
ho, snpid,  
heidi_outli  
er_flag=T,  
gwas_thresh  
=5e-8, sing  
le_heidi_th  
resh=0.01,  
multi_heidi  
_thresh=0.0  
1, nsnp_s_th  
resh=10, ld
```

```
_r2_thresh=  
0.05, ld_fd  
r_thresh=0.  
05, gsmr2_b  
eta=0)
```

Arguments

bxz

bxz_se

bzx_pval

bzy

bzy_se

ldrho

snpid

n_ref

heidi_outlier fla

gwas_thresh

nsnps_thresh

ld_r2_thresh

ld_fdr_thresh

gsmr2_beta

single_heidi_thro

multi_heidi_thres

Value

Estimate of
causative
effect of risk
factor on
disease (b_{xy}),
the
corresponding
standard error

(bxy_se), p-
value

(bxy_pval),

SNP index

(used_index),

SNPs with

missing

values, with

non-
significant p-
values and
those in LD.

Examples

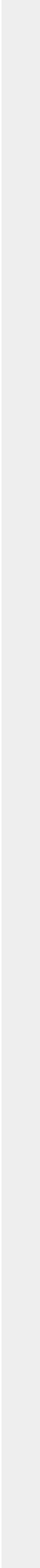
```
data("gsmr  
")  
gsmr_result
```



```
t = gsmr(g  
smr_data$b  
zx, gsmr_d  
ata$bzx_se  
, gsmr_dat  
a$bzx_pval  
, gsmr_dat  
a$bzy, gsm  
r_data$bzy  
_se, ldrho  
, gsmr_dat  
a$SNP . n r
```

```
ef, T, 5e-  
8, 0.01, 0  
.01, 10, 0  
.1, 0.05,  
0)
```

 **std_effect**



Standardization
of SNP
effect and
its
standard
error using
z-statistic,
allele
frequency



and sample
size

Usage

```
std_effect(  
  snp_freq, b  
  , se, n)
```

Arguments

snp_freq

vector,

allele

frequen

b

vector,
SNP
effects
risk fac

se

vector,
standar
errors c

n

vector,
SNP
sample
sizes fo
GWAS
the risk
factor

Value

Standardised
effect (b) and
standard error
(se)

Examples

```
data("gsmr
```

```
" )  
std_effect  
s = std_ef  
fect(gsmr_  
data$a1_fr  
eq, gsmr_d  
ata$bzx, g  
smr_data$b  
zx_se, gsm  
r_data$bzx  
_n)
```

